

1 **The therapeutic effect of atovaquone and clindamycin on the reduction of tissue cysts of**
2 **PRU strain of *Toxoplasma gondii***

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12
13 **Abstract**

14 Despite recent advances in the treatment of cerebral toxoplasmosis, monitoring parasite load and
15 treatment response is still challenging. In the present study, the effect of atovaquone (AT) and
16 clindamycin (CL) alone and in combination on chronic cerebral toxoplasmosis caused by PRU
17 strain of *Toxoplasma gondii* was investigated in BALB/c mice. BALB/c mice aged 6 to 8 weeks
18 were infected by intraperitoneal inoculation of *Toxoplasma gondii* strain PRU brain cysts. Then,
19 the mice were divided into five groups as follows: Group 1 included mice treated with 100 mg/kg
20 of atovaquone (AT), group 2 included mice treated with 400 mg/kg per day of clindamycin, group
21 3 included mice treated with combination (AT+CL), group 4 included untreated, infected mice as
22 a positive control (PC) and group 5 included untreated uninfected mice as negative control (NC).
23 After the completion of the treatment period, the effect of drugs in reducing or eliminating parasites
24 in the brain was checked by counting the number of brain cysts. The results showed that although
25 atovaquone and clindamycin did not completely remove the cysts from the brain tissue of mice,
26 they significantly reduced the number of tissue cysts in the brain tissue of the treated mice
27 compared to the untreated control group (PC) ($P < 0.0001$). Atovaquone had more anti-
28 toxoplasmic effect than clindamycin and the difference between the two drugs was completely
29 significant. In conclusion, considering the risks of infection with different strains of *T. gondii*, it is
30 necessary to emphasize the importance of developing effective therapeutic interventions for
31 toxoplasmosis.

32 **Key words:** *Toxoplasma gondii*, cerebral toxoplasmosis, PRU strain, atovaquone, clindamycin.

33
34 **1. Introduction**

35 Toxoplasmosis is a globally widespread infection, with particularly high rates in some areas. The
36 demand for new treatments or drugs for toxoplasmosis arise from several key reasons. Current

37 therapies, mainly pyrimethamine and sulfadiazine, often cause severe side effects and require close
38 monitoring of blood levels to prevent toxicity, particularly during prolonged use (1). Another
39 growing issue is the potential for *Toxoplasma gondii* to develop drug resistance, which may reduce
40 the efficacy of existing medications. Immunocompromised individuals—such as those with
41 HIV/AIDS, undergoing cancer therapy, or receiving organ transplants—face a greater risk of
42 severe toxoplasmosis, and available treatments may not adequately control infections in these
43 patients. Additionally, congenital toxoplasmosis (transmitted during pregnancy) can result in fetal
44 complications like neurological impairment, vision loss, and developmental delays. Better
45 therapeutic approaches could significantly improve outcomes for both mothers and infants (1).

46 Atovaquone is an antiprotozoal drug designed to prevent and treat specific protozoal infections. It
47 functions as a structural analog of coenzyme Q (ubiquinone), a critical element in mitochondrial
48 electron transport. By binding to cytochrome b, atovaquone disrupts the mitochondrial membrane
49 potential and interferes with pyrimidine synthesis in the parasite (2, 3), ultimately impairing energy
50 production and causing cell death.

51 As a standalone treatment, atovaquone is effective against *Toxoplasma gondii* and *Pneumocystis*
52 *jiroveci* infections, as well as malaria prevention (2, 3). In vitro studies demonstrate its potent
53 activity against tachyzoites at nanogram-per-milliliter concentrations (4, 5), though higher doses
54 are needed to eliminate bradyzoites within cysts (4). In mouse models of toxoplasmosis,
55 atovaquone showed efficacy alone (6), but its effectiveness improved when combined with other
56 agents, such as pyrimethamine, sulfadiazine (7), clindamycin (8), azithromycin (9), or
57 clarithromycin (5). An experimental intravenous formulation also demonstrated high efficacy in a
58 reactivated toxoplasmosis mouse model (10).

59 Clindamycin, a lincosamide-class antibiotic, targets both aerobic and anaerobic bacteria by
60 inhibiting protein synthesis. It binds to the bacterial 50S ribosomal subunit, preventing bacterial
61 growth. Beyond its antibacterial uses, clindamycin has also been employed—alone or in
62 combination—for toxoplasmosis treatment (11, 12).

63 The need for novel toxoplasmosis therapies remains urgent to overcome current treatment
64 limitations, safeguard high-risk populations, and reduce the global impact of this infection.
65 Continued research is vital to advancing therapeutic options and improving patient outcomes. To
66 address these gaps, this study systematically evaluates the anti-parasitic efficacy of atovaquone
67 both as monotherapy and in combination with clindamycin using a well-established Balb/c mouse
68 model infected with the clinically relevant PRU strain of *T. gondii*.

69 **Materials and Methods**

70 **1.1 Mice**

71 Female Balb/c mice (Razi Vaccine & Sera Institute, Karaj, Iran) weighing 20 to 25 g at the
72 beginning of each experiment were used. Mice were housed 4 to a cage and offered drinking water
73 ad libitum.

74 **1.2 Parasite**

75 The *T. gondii* PRU strain used in this study was provided by the Parasitology Department at
76 Mazandaran University of Medical Sciences. To establish chronic infection in BALB/c mice, the
77 parasite tachyzoites were maintained by serial intraperitoneal passage in laboratory mice. Three
78 chronically infected donor mice were euthanized via chloroform anesthesia and surface-sterilized
79 in 96% ethanol. Under a biosafety hood, skulls were aseptically opened, and brains were extracted.
80 Brain tissue was rinsed in sterile distilled water, and cysts were confirmed microscopically (400×
81 magnification). Tissue was gently homogenized to preserve cyst integrity. Homogenized brain
82 suspension was adjusted to 2 mL with sterile water and further dispersed using a 2.5 mL syringe.
83 Cysts were enumerated via Neubauer chamber to standardize the inoculum. Each mouse received
84 an intraperitoneal injection containing 20–25 cysts. Infected mice were ear-marked, randomized
85 into five groups, and housed individually.

86 **1.3 Drugs**

87 Clindamycin (CLI): Hydrochloride powder (Sepidaj Pharmaceutical Co., Iran) was administered
88 at 400 mg/kg/day. Atovaquone (ATO): Micronized powder (Hubei Vanz Pharm Co., China) was
89 given at 100 mg/kg/day. Doses were chosen based on prior studies demonstrating efficacy (13, 14)
90 as effective doses (400 mg CLI/kg/day and 100 mg ATO/kg/day). Suboptimal doses (Lower doses
91 of both drugs) were also tested to better evaluate combination therapy effects. Infrared (IR)
92 spectroscopy was performed at Tarbiat Modares University's Faculty of Medical Sciences to
93 confirm drug integrity. To control for drug side effects, separate groups of animals were given CLI
94 (400 mg/kg/day) and ATO (100 mg/kg/day) for 3 months, as well as ATO plus CLI (100 plus 400
95 mg/kg/day) for 2 months.

96 **1.4 Experimental design.**

97 Mice were intraperitoneally inoculated with 20-25 *T. gondii* cysts and randomly divided into
98 treatment groups (n=12 per group): 1- CLI monotherapy (400 mg/kg/day); 2- ATO monotherapy
99 (100 mg/kg/day) and 3- Combination therapy (ATO+CLI: 100+400 mg/kg/day). Two control
100 groups were included: 1- Negative control (NC): Uninfected mice and 2- Positive control (PC):
101 Infected, untreated mice. Following inoculation, mice were housed for 8 weeks to establish chronic
102 infection. Treatment began 24 hours post-inoculation and continued for 14 consecutive days.
103 Survival was monitored daily throughout the experiment. The complete study was replicated three
104 times with consistent results. Data presented represent pooled results from all replicates.

105 **1.5 Bioassay.**

106 Following the 14-day treatment period, mice were euthanized using chloroform anesthesia. Under
107 sterile biosafety cabinet conditions, mice were secured on a dissection tray, skulls were aseptically
108 opened using surgical tools (scalpel and forceps) and whole brains were extracted and rinsed with
109 sterile distilled water. Each brain was divided into two hemispheres, left hemisphere was stored at
110 -80°C in sterile microtubes for molecular analysis and right hemisphere was used for tissue cyst
111 quantification. Brain hemispheres designated for cyst counting were homogenized 20-25 cysts
112 from each sample were intraperitoneally inoculated into two naive mice per sample (1 mouse per
113 inoculation route).

114 **1.6 Statistical analysis.**

115 The cyst count data were analyzed using one-way ANOVA in GraphPad Prism software (version
116 X.X), with statistical significance set at $p < 0.05$. Post-hoc multiple comparisons were performed
117 when ANOVA indicated significant differences between groups.

118 **2. Results**

119
120 Following the 14-day treatment regimen, quantitative analysis revealed, neither atovaquone nor
121 clindamycin monotherapy achieved complete cyst eradication. All treatment groups showed
122 statistically significant reductions in brain cyst counts compared to untreated controls (PC) ($p <$
123 0.0001). The magnitude of cyst reduction varied by treatment regimen (see Table 1 and Figure 1)

124
125 **Table 1:** The number of cysts counted in the brains of different groups of mice after treatment
126 with atovaquone and clindamycin.
127

Groups	Drugs used	Cyst /g in brain	P- value*
1	Atovaquone (AT)	163.33 ± 29.52	2, 3, 4
2	Clindamycin (CL)	496 ± 41.85	1, 3, 4
3	AT + CL	1096.8 ± 84.82	1, 2, 4
4	No treatment for positive control (PC)	1600.8 ± 75.91	1, 2, 3

128 *significant differences in various groups based on One-way ANOVA test.

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129

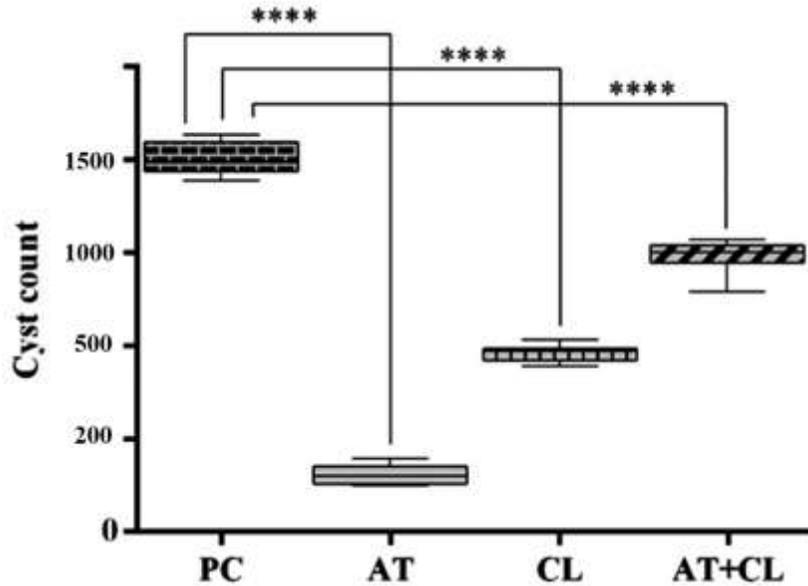


Fig. 1: The effect of treatment with Atovaquone, Clindamycin and combined drug (Atovaquone + Clindamycin) on the number of tissue cysts in the brain tissue of mice. PC: Positive Control (No treatment), AT: Atovaquone, CL: Clindamycin.

In addition, a significant variation in cyst counts was observed across all treatment groups ($p < 0.0001$). Atovaquone (ATO) monotherapy demonstrated superior efficacy, with significantly fewer cysts than Clindamycin (CLI) monotherapy ($p < 0.0001$) and combination therapy (ATO+CLI) ($p < 0.0001$). CLI monotherapy also showed reduced efficacy compared to the combination group ($p < 0.0001$) (Figure 1).

ATO exhibited significantly stronger anti-toxoplasmic activity than CLI. The ATO+CLI combination paradoxically showed lower efficacy than ATO alone, suggesting drug antagonistic interaction and interference against the PRU strain. No treatment-related toxicity (e.g., lethargy, weight loss) was observed during extended maintenance (2–3 months) in any drug-treated group.

4. Discussion

T. gondii exhibits limited species diversity but comprises three major strain types with distinct biological properties. Type I (e.g., RH, GT1) is highly virulent in mice, often lethal in acute infection models. Type II (e.g., PRU, ME49) is moderately virulent; preferentially establishes chronic infections in intermediate hosts (including humans) and type III (e.g., CEP) is attenuated virulence, primarily used for comparative studies (16). As a canonical Type II strain, PRU is widely employed in toxoplasmosis research due to its clinical relevance as a model for human chronic infection and cyst persistence. It is often used in laboratory studies to understand *T. gondii*

154 biology, host-parasite interactions, and the immune response. It helps researchers to investigate
155 pathogenesis, therapeutic development and vaccine design (15).

156 Toxoplasmosis represents a major public health concern due to its severe consequences for
157 pregnant women and immunocompromised individuals. As awareness of its health impacts
158 increases, so does the need for more effective and accessible treatments. Recent advances in our
159 understanding of *T. gondii* biology have uncovered promising new drug targets, including essential
160 metabolic pathways, critical signaling mechanisms, and vulnerable stages of the parasite's lifecycle
161 that could be exploited for therapeutic development. Novel drug compounds or combination
162 therapies may provide improved treatment outcomes through enhanced efficacy, reduced toxicity,
163 or innovative mechanisms of action. The development of such treatments could significantly
164 reduce cases of severe toxoplasmosis, thereby decreasing healthcare burdens and improving
165 quality of life for affected populations (16).

166 The management of toxoplasmosis varies based on patient age, immune status, and clinical
167 presentation. While most available medications effectively target the tachyzoite stage, they
168 demonstrate limited activity against persistent tissue cysts. Naphthoquinone derivatives such as
169 atovaquone represent an exception, showing modest cysticidal activity in experimental models,
170 though their clinical use has primarily been documented in HIV/AIDS patients (17). The structural
171 complexity of the cyst wall presents a significant therapeutic challenge.

172 In clinical practice, the combination of pyrimethamine with either sulfadiazine or clindamycin
173 remains the gold standard for both acute treatment and secondary prophylaxis (17,18). However,
174 treatment-limiting toxicity frequently necessitates alternative approaches in a substantial
175 proportion of patients.

176 Our experimental findings demonstrate that while neither atovaquone nor clindamycin
177 monotherapy achieved complete cyst eradication in the *T. gondii* PRU strain model, both agents
178 significantly reduced cerebral cyst burden compared to untreated controls. Notably, atovaquone
179 exhibited superior anti-parasitic efficacy relative to clindamycin, with statistically significant
180 differences in cyst reduction between the two treatment modalities.

181 In the study by Moshkani and Dalimi (2000), mice infected with *T. gondii* RH strain tachyzoites
182 were treated with atovaquone and azithromycin, either alone or in combination. The results showed
183 dose-dependent survival rates with atovaquone monotherapy: 8%, 17%, and 25% of mice survived
184 at doses of 20, 50, and 100 mg/kg/day, respectively. While azithromycin failed to eradicate the
185 parasite from either brain or visceral tissues, atovaquone demonstrated complete clearance of
186 visceral infection at all tested doses (20-100 mg/kg/day) and eliminated brain infection at the
187 highest dose (100 mg/kg/day). Notably, combination therapy with atovaquone and azithromycin
188 showed neither synergistic nor additive effects and failed to achieve complete parasite eradication
189 in any tissue compartment (9).

190 Djurković-Djaković et al. (1999) evaluated the efficacy of clindamycin (CLI) combined with
191 atovaquone (ATO) in a murine model of acute toxoplasmosis. Swiss Webster mice were
192 intraperitoneally infected with either 10^2 or 10^4 tachyzoites of the *T. gondii* RH strain and received
193 oral treatment with each drug alone or in combination for 14 days starting from day 1 post-

194 infection, with survival monitored over 7 weeks. In mice infected with 10^2 parasites, the drug
195 combination significantly improved survival compared to ATO monotherapy, though it provided
196 no additional benefit over CLI alone, which showed strong efficacy. For the higher inoculum (10^4
197 parasites), the combination therapy outperformed ATO alone at both low and high doses but again
198 showed no advantage over CLI monotherapy (8).

199 In a follow-up study in 2002, the same research group examined this drug combination against the
200 ME49 strain in Swiss-Webster mice orally infected with 10 or 20 cysts. Treatment with ATO (5–
201 100 mg/kg/day) and CLI (25–400 mg/kg/day), either alone or combined, was administered for 2–
202 4 weeks. In acute infection, all treatments significantly enhanced survival and reduced brain cyst
203 burden, with ATO-containing regimens (both monotherapy and combination) showing superior
204 cyst reduction compared to CLI alone. For chronic infection, only the combination therapy
205 achieved a significant decrease in cyst burden when assessed 2 weeks post-treatment (19).

206 Several studies have evaluated atovaquone's effectiveness against toxoplasmosis in various animal
207 models. In hamsters with acute acquired toxoplasma retinochoroiditis, systemic atovaquone
208 monotherapy demonstrated comparable efficacy to standard regimens (pyrimethamine-
209 sulfadiazine, clindamycin, and spiramycin) in reducing *Toxoplasma* brain cyst burden during acute
210 infection. Notably, atovaquone also significantly decreased cyst numbers in chronic infection (20).

211 Formulation advancements have enhanced atovaquone's therapeutic potential. Azami et al. (2018)
212 developed an atovaquone nanoemulsion that showed improved bioavailability and tissue
213 distribution in mice infected with both RH and Tehran *T. gondii* strains. This formulation increased
214 survival time while reducing parasitemia, brain cyst count, and cyst size (21). More recently,
215 Goudarzi et al. (2024) demonstrated that atovaquone-loaded exosomes (EXO-ATQ) achieved
216 97.3% cyst reduction in chronic infection (Tehran strain) and showed enhanced efficacy against
217 tachyzoite proliferation in vitro compared to conventional atovaquone suspension (22).

218 Beyond treatment, atovaquone shows promise for toxoplasmosis prophylaxis in transplant
219 recipients, though its safety and efficacy in this specific population require further investigation
220 (4, 23).

221 Collective research findings position atovaquone as a promising therapeutic candidate against *T.*
222 *gondii*, demonstrating superior cyst-reducing efficacy compared to conventional therapies. Its
223 potential integration into future treatment protocols is further supported by evidence that
224 combination regimens can significantly enhance therapeutic outcomes. These advances are
225 particularly critical given toxoplasmosis' growing public health burden, especially among
226 immunocompromised individuals and pregnant women, underscoring the urgent need for
227 continued investigation into parasite biology and host-pathogen dynamics.

228 While pyrimethamine-sulfadiazine remains the current standard of care, its limitations—including
229 treatment-limiting toxicity and incomplete cyst eradication—necessitate the development of safer,
230 more effective alternatives. A multidisciplinary approach combining parasitology, drug
231 development, and clinical research will be essential to advance next-generation therapies. Such
232 innovations must address the full spectrum of toxoplasmosis management, from acute infection to
233 chronic cyst clearance, ultimately improving therapeutic efficacy and patient quality of life.

234

235 **Certificate of medical ethics**

236 This study was approved by the ethics committee of Tarbiat Modares University with code number
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238

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243 **Authors' contributions**

244 Study concept and design: N.Z. and A.D.

245 Acquisition of data: N.Z.

246 Analysis and interpretation of data: A.D.

247 Drafting of the manuscript: A.D. and N.Z.

248 Critical revision of the manuscript for important intellectual content: A.D.

249 Statistical analysis: A.D.

250 Administrative, technical, and material support: A.D.

251 **Conflict of interest**

252

253 The authors have declared no conflicts of interest.

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259

260 **Data Availability**

261 The data used to support the findings of this study are available from the corresponding author
262 upon reasonable request.

263

264 **References**

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