

1 **Molecular and bioassay examination of *Neospora caninum* infection in bovine aborted**
2 **fetuses in Khorasan Razavi province, Iran.**

3 **Abstract**

4 *Neospora caninum* plays a significant role in causing abortion and reproductive failure in
5 dairy cattle. The majority of neosporosis-related abortions take place during the 5–6 months
6 of gestation. Fetal death in the uterus, resorption, mummification, autolysis, stillbirth, birth
7 with clinical symptoms, or being born clinically healthy but with chronic infection are all
8 possible outcomes. The objective of the study was to identify *N. caninum* infection in aborted
9 bovine fetuses through molecular analysis and mouse bioassay testing. From 2019 to 2022,
10 121 bovine aborted fetuses were collected from dairy farms in Khorasan Razavi province.
11 The fetal brain samples were screened for detection of the parasite DNA by polymerase chain
12 reaction assay (PCR). In addition, a portion of PCR-positive brain tissue was homogenized
13 and inoculated into the peritoneum of five BALB/c mice. All mice were sacrificed six weeks
14 post infection and examined using serology, microscopic, and PCR methods. If the mice's
15 brain samples were PCR positive, the mouse bioassay test was repeated two times. The *N.*
16 *caninum* DNA was detected in 19.8% of brain samples in bovine aborted fetuses. Among
17 PCR-positive brain samples, only ten samples were suitable for mouse bioassay examination.
18 All inoculated mice were seronegative without clinical signs after three times bioassays,
19 although the brain samples of three mice groups were PCR-positive after repeated bioassays.
20 The PCR results showed a moderate frequency of *N.caninum* infection in aborted bovine
21 fetuses. Furthermore, the isolates obtained in this study had low pathogenicity in BALB/c
22 mice. It seems the isolates belong to an avirulent strain

23 **Keywords:** *Neospora*; Abortion; Cattle; PCR; Bioassay examination;

- ۲۴
- ۲۵ بررسی مولکولی و زیست‌سنجی نیوسپورا کانینوم در جنین‌های سقط شده گاوی در منطقه مشهد، استان
- ۲۶ خراسان رضوی، ایران.
- ۲۷ نیوسپورا کانینوم نقش بسزایی در ایجاد سقط جنین و نارسایی تولید مثل در گاوهای شیری دارد. اکثر سقط‌های مربوط به
- ۲۸ نئوسپوروزیس در طی ۵ تا ۶ ماه بارداری اتفاق می‌افتد. مرگ جنین در رحم، مومیایی شدن، اتولیز، مرده‌زایی، تولد با علائم
- ۲۹ بالینی یا تولد از نظر بالینی سالم اما با عفونت مزمن، همگی پیامدهای احتمالی این آلودگی هستند. هدف از این مطالعه تعیین
- ۳۰ آلودگی نیوسپورا در جنین‌های گاو سقط‌شده از طریق آنالیز مولکولی و تست زیست‌سنجی موش بود. از سال ۲۰۱۹ تا
- ۳۱ ۲۰۲۲، تعداد ۱۲۱ جنین سقط شده گاوی از گاوداری‌های استان خراسان رضوی جمع‌آوری شد. نمونه‌های مغز جنین برای
- ۳۲ یافتن DNA انگل با روش واکنش زنجیره‌ای پلیمرز غربالگری شدند. علاوه بر این، بخشی از بافت مغزی جنین‌ها را که در
- ۳۳ آزمایش واکنش زنجیره‌ای پلیمرز مثبت شده بودند، پس از طی مراحل همگن‌سازی به روش داخل صفاق به پنج موش
- ۳۴ BALB/c تلقیح شدند. در پایان تجربه، همه موش‌ها شش هفته پس از آلودگی به روش انسانی کشته شده و نمونه‌های
- ۳۵ خونی و بافتی آنها با روش‌های سرولوژی، میکروسکوپی و واکنش زنجیره‌ای پلیمرز مورد بررسی قرار گرفتند. چنانچه
- ۳۶ نمونه‌های مغز موش‌ها با آزمایش واکنش زنجیره‌ای پلیمرز مثبت نشان می‌دادند، آزمایش زیست‌سنجی موش جهت
- ۳۷ جدا سازی بهتر دو باره تکرار می‌گردید. در این بررسی در ۱۹/۸ درصد نمونه‌های مغز در جنین‌های سقط شده گاو DNA
- ۳۸ نیوسپورا شناسایی شد. از بین نمونه‌های مغزی PCR مثبت، تنها ۱۰ نمونه برای بررسی زیست‌سنجی در موش‌ها مناسب
- ۳۹ بودند. همه موش‌های تلقیح شده پس از سه بار آزمایش زیست‌سنجی فاقد علائم بالینی بوده و در آزمایش سرولوژی نیز
- ۴۰ نتیجه منفی نشان دادند، در صورتیکه آزمایش PCR در تعدادی از نمونه‌های مغزی دو گروه از موش‌های تلقیح شده پس از
- ۴۱ دو آزمایش زیستی سنجی مکرر مثبت بودند. نتایج این مطالعه نشان داد که درصد قابل توجهی از جنین‌های سقط شده گاو

۴۲ آلوده به نیوسپورا بودند . علاوه بر این ، این جدایه ها در موش BALB/C بیماری زایی پایینی داشتند، بنظر می رسد این

۴۳ جدایه ها متعلق به سویه های غیر حاد نیوسپورا باشند.

۴۴ **کلمات کلیدی:** نیوسپورا، سقط جنین، گاو، واکنش زنجیره ای پلیمرز، بررسی زیست‌سنجی

۴۵

۴۶ **1. Introduction**

۴۷ *Neospora caninum* is recognized as the primary cause of abortion in cattle worldwide (1).

۴۸ There are two ways that *N. caninum* can be transmitted to cattle. The main method of

۴۹ infection is typically referred to as vertical, congenital, or endogenous transmission. This

۵۰ occurs when the tachyzoites of cysts in the dam's tissues become active and transform into

۵۱ tachyzoites, which then travel through the placenta and into the fetus. The secondary method

۵۲ of infection is known as horizontal or post-natal transmission. This happens when pregnant

۵۳ dairy cattle consume sporulated *N. caninum* oocysts, and the sporozoites transform into

۵۴ tachyzoites, likely spreading through the circulation in cells of the mononuclear phagocytic

۵۵ system and potentially infecting the fetus through transplacental transmission (2). The

۵۶ endogenous (vertical) transmission could occur in approximately up to 95% of infected dairy

۵۷ cattle (1,3). The high seroprevalence of *Neospora* infection has been shown in dairy cattle in

۵۸ Iran (4,5,6,7). Hence, it was found that *N. caninum* infection also plays a role in causing

۵۹ abortion in dairy cattle (8,9). Although there is a high prevalence of *N. caninum* infection in

۶۰ dairy cattle in Iran, only one *N. caninum* isolate was recovered from a bovine fetus that was

۶۱ aborted (10). The present study aimed to detect the frequency of *N. caninum* infection in

۶۲ aborted bovine fetuses and evaluate the pathogenicity of *N. caninum* in BALB/c mice

۶۳

٦٤ **2. Materials and Methods**

٦٥ The research was conducted in the northeastern province of Khorasan Razavi in Iran,
٦٦ covering an area of over 127,000 km² and located at coordinates 33°30'-37°41' N; 56°19'-
٦٧ 61°18' E. This province falls within the northern temperature zone and experiences a semi-
٦٨ arid climate characterized by cold winters and moderate summers. There are approximately
٦٩ 25000 cattle distributed across 110 dairy farms in this region, with herd sizes ranging from 30
٧٠ to 2000 cattle varying between farms. The predominant cattle breed in the area is Holstein-
٧١ Friesian.

٧٢ **2.1. Sample collection**

٧٣ From 2022 to 2023, 121 aborted bovine fetuses were collected from different areas of the
٧٤ province. Then, the fetuses were necropsied, and collected the brain tissue for molecular and
٧٥ bioassay examination

٧٦ **2.2. DNA extraction and PCR**

٧٧ Samples were used to extract genomic DNA with the MBST Genomic DNA kit (Molecular
٧٨ and Biological Transmission Systems, Tehran, Iran) following the manufacturer's
٧٩ instructions. After that, we conducted a PCR assay to identify the *N.caninum* gene, following
٨٠ the previously described method by Müller et al 2002(11).

٨١ The simple PCR reaction was performed in a 25µl mixture containing 2µl of total DNA, 10µl
٨٢ of commercial premix master mix (Parstous Mashhad), 1µl of each primer, and 11µl of
٨٣ nuclease-free water in a thermocycler. The cycling process began with an initial denaturation
٨٤ step at 95 °C for 5 minutes, followed by 40 cycles of 94 °C for 1 minute, 63 °C for 1 minute,
٨٥ and 74 °C for 3.5 minutes, and concluded with a final extension step at 74 °C for 10 minutes.

٨٦ The Oligonucleotide primers used were NP21plus
٨٧ (5'CCCAGTGCGTCCAATCCTGTAAC3') and Np6plus
٨٨ (5'CTCGCCAGTCAACCTACGTCTTCT3').

٨٩ **2.3. Bioassay in mice**

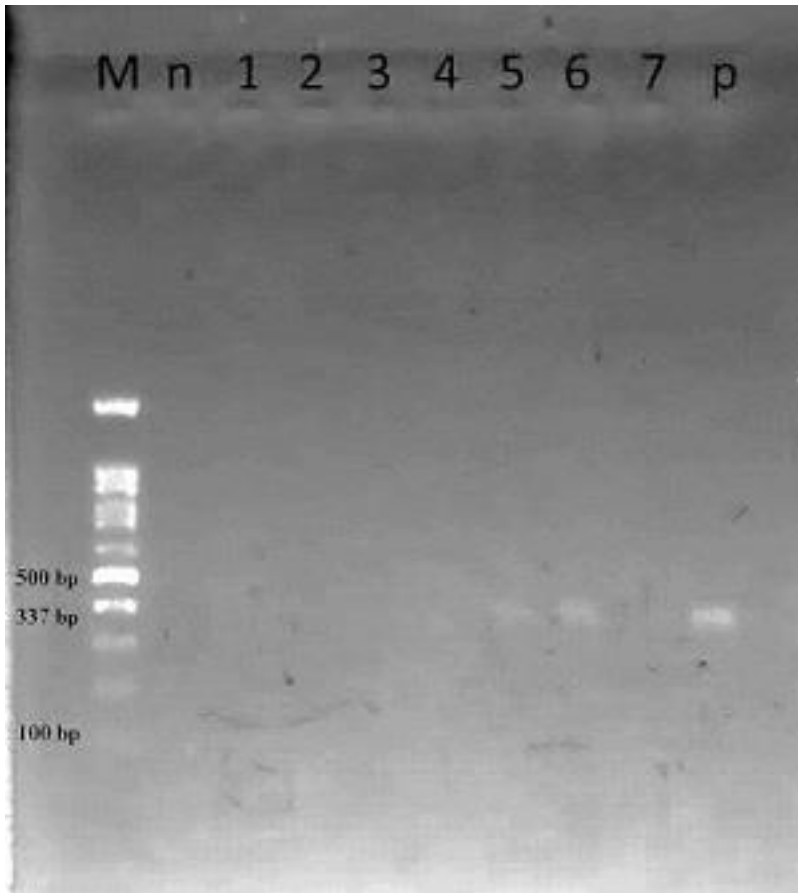
٩٠ Six to eight-week-old female BALB/c mice were acquired from the Razi Vaccine and Serum
٩١ Research Institute (Mashhad Branch). The mice were kept in plastic box cages in groups of 5
٩٢ and given rodent feed and water ad libitum under standard conditions. A total of 100 grams of
٩٣ brains from PCR-positive aborted fetuses were homogenized in 500 ml of 0.85% NaCl
٩٤ solution (saline) containing antibiotics (100 IU/ml penicillin and 745 IU/ml streptomycin)
٩٥ and then homogenized using an electrical mixer. The mixture was subsequently filtered
٩٦ through two layers of gauze. After an incubation period of three hours at room temperature,
٩٧ the samples were centrifuged at 1500 g for five minutes, and 5 ml of the homogenate deposit
٩٨ was administered intraperitoneally to 5 BALB/c mice (1 ml per mouse).

٩٩ The mice were observed daily for any clinical signs indicating neosporosis. Blood samples
١٠٠ were obtained from the mice's tails six weeks after they were inoculated, and their serum
١٠١ samples were analyzed using an Elisa kit (ID screen ® *N. caninum* indirect Multi-species, ID.
١٠٢ vet, Montpellier, France) to detect *Neospora* antibodies. All the mice that were inoculated
١٠٣ were euthanized 42 days after being infected using chloroform inhalation. Brain impression
١٠٤ smears were prepared and examined under a microscope to detect cysts. A portion of the
١٠٥ brain tissue was tested for *N.caninum* DNA using PCR. The PCR-positive brain samples from
١٠٦ each group of mice were combined and then inoculated intraperitoneally into five BALB/C
١٠٧ mice. These inoculated mice were observed daily and then euthanized seven weeks after
١٠٨ being inoculated. During the necropsy, blood and brain samples were collected and analyzed

109 using the serology and PCR methods mentioned earlier. If the brain samples from the mice
110 tested positive via PCR, the mice bioassay examination was repeated to once again find a
111 viable cyst.

112 **3. Results**

113 In the present study, *N.caninum* DNA was detected in 19.8% (24/121) of brain samples of
114 bovine aborted fetuses (Fig1). Among 24 PCR-positive brain samples, only 10 samples were
115 suitable for mouse bioassay examination. In the first bioassay round, All inoculated mice in
116 ten groups were normal without clinical signs and the serology and microscopy results were
117 also negative after 42 days post-infection, but, *N.caninum* -DNA were detected in five brain
118 samples of two mice groups by PCR. Similar results were obtained after two rounds of mice
119 bioassay examination in BALB/C mice that were inoculated with PCR-positive mice brains
120 of infected groups. (Table 1).



۱۲۱

۱۲۲

Fig.1. PCR amplification products of *N. caninum* in brain samples Lanes: M: molecular

۱۲۳

weight marker (between 1000 and 100bp); p: positive control (*Neospora tachyzoites*; 337

۱۲۴

bp); n: negative control; 5, 6: positive samples.

۱۲۵

۱۲۶

۱۲۷

۱۲۸

۱۲۹

۱۳۰

۱۳۱

Table1- The results of first, second and third round of mouse bioassay examination on PCR -positive bovine and mice brain samples.

Results	First round			Second round			Third round		
	bovine aborted fetuses (homogenized brain)			Mice			Mice		
Group	ELISA	Microscopy	PCR	ELISA	Microscopy	PCR	ELISA	Microscopy	PCR
(5 mice)									
1	N	N	N	nd	nd	nd	nd	nd	nd
2	N	N	N	nd	nd	nd	nd	nd	nd
3	N	N	P	N	N	P	N	N	P
4	N	N	P	N	N	N	nd	nd	nd
5	N	N	N	nd	nd	nd	nd	nd	nd
6	N	N	N	nd	nd	nd	nd	nd	nd
7	N	N	P	N	N	P	N	N	P
8	N	N	N	nd	nd	nd	nd	nd	nd
9	N	N	P	N	N	N	nd	nd	nd
10	N	N	P	N	N	P	N	N	P

۱۳۳ P=Positive, N=Negative, nd= not done

۱۳۴

Preprint

۱۳۵ **4. Discussion**

۱۳۶
۱۳۷ In this research, *N. caninum* DNA was identified in 19.8% (24 out of 121) of brain samples
۱۳۸ taken from aborted bovine fetuses using the PCR method. The prevalence of *N. caninum*
۱۳۹ infection in aborted bovine fetuses across various provinces in Iran has been reported to range
۱۴۰ from 12% to 67%, as determined by PCR techniques (Table 2). A meta-analysis indicated that
۱۴۱ the prevalence of *N. caninum* in aborted fetuses was greater in studies with fewer than 50
۱۴۲ samples compared to those with more than 50 samples (23). The author concluded that the
۱۴۳ pooled estimate for the subgroup with a sample size of 50 or more could provide a more
۱۴۴ accurate, conservative, and reliable representation of the overall infection rates in aborted
۱۴۵ bovine fetuses in Iran (23).

۱۴۶
۱۴۷
۱۴۸
۱۴۹
۱۵۰
۱۵۱
۱۵۲
۱۵۳
۱۵۴
۱۵۵
۱۵۶
۱۵۷
۱۵۸

Table2- List of different studies frequency of *N. caninum* infection in brain tissue of aborted bovine fetuses in different areas of Iran

province	Study Year	Method	Number examined	Number infected	Frequency	References
Khorasan Razavi	2007	PCR	6	4	67%	(12)
Khorasan Razavi	2007	PCR, IHC	100	13	13%	(8)
Khorasan Razavi	2010	PCR	151	18	12%	(13)
Khorasan Razavi	2013	PCR	200	23	12%	(14)
East Azerbaijan	2013	PCR	14	6	43%	(15)
Charhar mahal & Bakhtiari	2013	PCR	100	11	11%	(16)
Tehran	2014	PCR	16	12	75%	(17)
Qazvin	2014	PCR	128	39	31%	(18)
East azerbaijan	2018	PCR	82	34	41%	(19)
Markazi	2018	PCR	38	10	26%	(20)
Mazandaran	2019	PCR	9	2	22%	(21)
Mazandaran	2021	PCR	78	16	20.5%	(22)

161 The brain tissue of bovine aborted fetuses is the main source of *N.caninum* isolation (24) but
162 some studies showed that most *N.caninum* cysts in brain tissue were probably non-viable due
163 to autolysis effect (25, 26). In this study, many positive brain samples were autolyzed after
164 abortion and only ten PCR-positive brain samples were suitable for bioassay examination. All
165 inoculated mice in ten bioassay groups showed no clinical signs and no *Neospora* cysts in
166 mice brain tissue, but *N.caninum* DNA was detected in the brain samples of five mice in three
167 bioassay groups. The same results were obtained after conducting two repetitive mouse
168 bioassay examinations with PCR-positive brain samples of mice. It seems that these
169 *N.caninum* isolates from aborted bovine fetuses were avirulent strains in BALB/C mice. So
170 far, virulent to avirulent strains of *N. caninum* have been reported by mice bioassay method
171 (24, 27).

172 The BALB/C models have been used to determine of pathogenicity of *N. caninum* isolates in
173 bovine aborted fetuses. Some isolates had low virulence and were shown no clinical
174 symptoms and no detectable *N.caninum* cysts in mice brains (28-31).

175 The PCR results indicated the presence of *N.caninum* infection in the brain samples of
176 aborted bovine fetuses and in two groups of inoculated mice. However, it remains unclear
177 why no antibodies against *N. caninum* were found in inoculated mice after three rounds of
178 bioassays. Some studies also reported seronegative results for *Toxoplasma gondii* or
179 *N.caninum* infection in different laboratory animals when inoculated with infected mice or rat
180 brain tissue (32,33). They suggested that the lack of clinical signs and detectable antibodies
181 against to *T.gondii* or *N.caninum* infection may be due to the rapid death of parasites in the
182 neural tissue of mice, but had DNA intact present. The duration between the extraction and
183 analysis of brain tissue might have been excessive to support the viable tissue cysts in the

184 brain tissue (33). To sum up, this research demonstrated the existence of *N.caninum* infection
185 in brain-aborted bovine fetuses in the Mashhad region. The result strongly suggests the high
186 frequency endogenous transmission among dairy cattle in Iran. In addition, the bioassay
187 examination also provides further evidence that *N.caninum* isolates might be an avirulent
188 strain and needs to more investigation.

189

190 **Declarations and statements**

191 **Funding:**

192 This study was supported by Grant 3/45901 from the Vice President Research and
193 Technology of Veterinary Medicine, Ferdowsi University of Mashhad, Iran.

194 **Conflict of interests:**

195 The authors declare no conflict of interest.

196 **Data availability:**

197 The datasets generated during and/or analyzed during the current study are available from the
198 corresponding author upon reasonable request.

199 **Ethics and animal experimentation**

200 The mice were housed and maintained in the animal care facility at Ferdowsi University of
201 Mashhad. All animal experiments were performed in strict accordance with the guidelines
202 approved by the Animal Ethics Committee of our faculty IR.UM.REC.1399.063.

203 **Author contribution**

204 Study concept and design: G.R.R. Conduction: G.R.R. Molecular, Serological and Bioassay
205 examination:A.K., M.S., Formal analysis and investigation: G.R.R.,A.K., M.S., Writing -
206 original draft preparation: G.R.R., Funding acquisition: G.R.R.

۲۰۷ **Consent to participate:**

۲۰۸ Not applicable

۲۰۹ **Consent for publication:**

۲۱۰ Not applicable

۲۱۱ **Acknowledgments:**

۲۱۲ We would like to thank Dr Nargess Khleghnia and Dr Darya Fazael for helping to take
۲۱۳ samples from the submitted bovine aborted fetuses to the Excellence Research Center for
۲۱۴ Ruminantal Abortion and Neonatal Mortality in Mashhad area.

۲۱۵

۲۱۶ **References**

- ۲۱۷ 1. Dubey JP, Schares G, Ortega-Mora L.M. Epidemiology and Control of neosporosis and
۲۱۸ *Neospora caninum*. Clin Microbiol Rev. 2007; 20, 323–367.
- ۲۱۹ 2. Conraths J, Gottstein B, Neosporosis: General considerations. In: Ortega-Mora LM.,
۲۲۰ Gottstein B, Conraths FJ, Buxton D, editors. Protozoal abortion in farm ruminants, CAB
۲۲۱ International, Wallingford. 2007, p. 42-45.
- ۲۲۲ 3. Wouda W, Biology, Transmission, and Clinical signs in Ortega-Mora LM., Gottstein B.,
۲۲۳ Conraths FJ., Buxton D, editors. Protozoal abortion in farm ruminants. CAB International,
۲۲۴ Wallingford; 2007. p. 46-53.
- ۲۲۵ 4.Hajikolaei MH, Hamidinejat H, Ghorbanpoor M, Goraninejad S. Serological study of
۲۲۶ *Neospora caninum* infection in cattle from Ahvaz area, Iran. 2008 Oct; 2(2) :63-66.

- ۲۲۷ 5. Hadadzadeh HR, Shayan P, Vojgani M, Bolorchi M. Serological study of *Neospora*
۲۲۸ *caninum* in pregnant dairy cattle in Tehran, Iran. Iranian Journal of Veterinary Medicine.
۲۲۹ 2010 Jun; 4(2):113-116.
- ۲۳۰ 6. Ansari-Lari M. Bovine neosporosis in Iran: a systematic review and meta-analysis.
۲۳۱ Preventive Veterinary Medicine. 2020 Mar 1; 176:104913.5.
- ۲۳۲ 7. Gharekhani J, Yakhchali M, Berahmat R. *Neospora caninum* infection in Iran (2004–
۲۳۳ 2020): A review. Journal of Parasitic Diseases. 2020 Dec; 44(4):671-86.
- ۲۳۴ 8. Razmi GR, Maleki M, Farzaneh N, Talebkhan Garoussi M, Fallah AH. The first report of
۲۳۵ *Neospora caninum*-associated bovine abortion in the Mashhad area, Iran. Parasitol Res. 2007;
۲۳۶ 100:755-757.
- ۲۳۷ 9. Salehi N, Hadadzadeh H, Ashrafi Helen, J, Shayan P, Sadrebazzaz A. Molecular and
۲۳۸ pathological study of bovine aborted fetuses and placenta from *Neospora caninum* infected
۲۳۹ dairy cattle. Iranian Journal of Parasitology. 2009; 4(3): 40-51.
- ۲۴۰ 10. Salehi N, Haddadzadeh H, Shayan P, Koochi MK. Isolation of *Neospora caninum* from an
۲۴۱ aborted fetus of seropositive cattle in Iran. Veterinarski Arhiv. 2012, 82 (6), 545-553.
- ۲۴۲ 11. Müller N, Zimmermann V, Hentrich B, Gottstein B. Diagnosis of *Neospora caninum* and
۲۴۳ *Toxoplasma gondii* infection by PCR and DNA hybridization immunoassay. J Clin
۲۴۴ Microbiol. 1996; 34: 2850–2852.
- ۲۴۵ 12. Sadrebazzaz A, Habibi G, Haddadzadeh H, and Ashrafi J. Evaluation of bovine abortion
۲۴۶ associated with *Neospora caninum* by different diagnostic techniques in Mashhad, Iran.
۲۴۷ Parasitology research. 2007 May; 100:1257-60.
- ۲۴۸ 13. Razmi GR, Zarea H, Naseri Z. A survey of *Neospora caninum*-associated bovine abortion
۲۴۹ in large dairy farms of Mashhad, Iran. Parasitology research. 2010 May; 106:1419-23.

- ۲۵۰ 14. Razmi GR, Zarae H, Nourbakhsh MF, Naseri Z. Estimating the rate of transplacental
۲۵۱ transmission of *Neospora caninum* to aborted fetuses in seropositive dams in Mashhad area,
۲۵۲ Iran. Iranian Journal of Veterinary Medicine.2013; 7: 253-256.
- ۲۵۳ 15. Nematollahi A, Moghaddam GH, Jaafari R, Helan JA, Norouzi M. Study on outbreak of
۲۵۴ *Neospora caninum*-associated abortion in dairy cows in Tabriz (Iran) by serological,
۲۵۵ molecular and histopathologic methods. Asian Pacific Journal of Tropical Medicine. 2013
۲۵۶ Dec 1; 6(12):942-6.
- ۲۵۷ 16- Rafati N, Jaafarian M. The determination of prevalence of *Neospora caninum* in aborted
۲۵۸ fetuses in dairy cattle of Shahrekord area, Chahar Mahal Bakhtiari province, by Nested-PCR.
۲۵۹ Journal of Veterinary Laboratory Research. 2014 May 22; 6(1):45-50.
- ۲۶۰ 17. Salehi N, Gottstein B, Haddadzadeh HR. Genetic diversity of bovine *Neospora caninum*
۲۶۱ determined by microsatellite markers. Parasitology international. 2015 Oct 1; 64(5):357-61.
- ۲۶۲ 18. Kaveh AA, Merat E, Samani S, Danandeh R, Soltannezhad S. Infectious causes of bovine
۲۶۳ abortion in Qazvin Province, Iran. Archives of Razi Institute. 2017 Dec 1; 72(4):225-30.
- ۲۶۴ 19. Hosseini A, Merat E, Samani S, Nezhad SS, Danandeh R. Comparison of *Neospora*
۲۶۵ *caninum* infected tissues in aborted fetal bovine by PCR. Journal of Veterinary Research.
۲۶۶ 2018, 73: 377-382.
- ۲۶۷ 20. Khani M, Arabkhazaeli F, Hosseini SD, Shayan P. Molecular detection of *Neospora*
۲۶۸ *caninum* in aborted fetuses of cattle farms in Arak. Journal of Veterinary Research. 2019; 73:
۲۶۹ 457-463.
- ۲۷۰ 21. Amouei A, Sharif M, Sarvi S, Nejad RB, Aghayan SA, Hashemi-Soteh MB, Mizani A,
۲۷۱ Hosseini SA, Gholami S, Sadeghi A, Sarafrazi M. Etiology of livestock fetal mortality in
۲۷۲ Mazandaran province, Iran. PeerJ. 2019 Jan 18; 6: e5920.

22. Salehi B, Amouei A, Dodangeh S, Daryani A, Sarvi S, Safari-Kharyeki MR, Salehi S, Hosseini SA, Hosseini Z. Molecular identification of *Neospora caninum* infection in aborted fetuses of sheep, cattle, and goats in Mazandaran Province, Northern Iran. Iranian Journal of Parasitology. 2021 Jul; 16(3):483.
23. Ansari-Lari M. *Neospora caninum* in aborted bovine fetuses in Iran: a systematic review and meta-analysis. Annals of Parasitology. 2021; 67(3):354-366
24. Al-Qassab SE, Reichel MP, Ellis JT. On the biological and genetic diversity in *Neospora caninum* Diversity. 2010 Mar 22; 2(3):411-38.
25. Calarco L, Barratt J, Ellis J. Genome wide identification of mutational hotspots in the apicomplexan parasite *Neospora caninum* and the implications for virulence. Genome biology and evolution. 2018 Sep; 10(9):2417-31
26. Barr BC, Anderson ML, Blanchard PC, Daft BM, Kinde H, Conrad PA. Bovine fetal encephalitis and myocarditis associated with protozoal infections. Veterinary Pathology. 1990 Sep; 27(5):354-61.
27. Conrad PA, Barr BC, Sverlow KW, Anderson M, Daft B, Kinde H, Dubey JP, Munson L, Ardans A. In vitro isolation and characterization of a *Neospora* sp. from aborted bovine fetuses. Parasitology. 1993 Apr; 106(3):239-49.
28. Lindsay DS, Lenz SD, Cole RA, Dubey JP, Blagburn BL. Mouse model for central nervous system *Neospora caninum* infections. The Journal of parasitology. 1995 Apr 1:313-5.
29. Miller CM, Quinn HE, Windsor PA, Ellis JT. Characterization of the first Australian isolate of *Neospora caninum* from cattle. Australian Veterinary Journal. 2002 Oct; 80(10):620-5.

296 30. Rojo-Montejo S, Collantes-Fernández E, Regidor-Cerrillo J, Álvarez-García G, Marugan-
297 Hernández V, Pedraza-Díaz S, Blanco-Murcia J, Prenafeta A, Ortega-Mora LM. Isolation and
298 characterization of a bovine isolate of *Neospora caninum* with low virulence. Veterinary
299 parasitology. 2009 Jan 22; 159(1):7-16.

300 31. Dittrich, R.L., Regidor-Cerrillo, J., Ortega-Mora, L.M., de Oliveira Koch, M., Busch,
301 A.P.B., Gonçalves, K.A. and Cruz, A.A. Isolation of *Neospora caninum* from kidney and
302 brain of a bovine fetus and molecular characterization in Brazil. Experimental Parasitology,
303 2018; 185: 10-16.

304 32- Dubey JP, Shen SK, Kwok OC, Thulliez P. Toxoplasmosis in rats (*Rattus norvegicus*):
305 congenital transmission to first- and second-generation offspring and isolation of *Toxoplasma*
306 *gondii* from seronegative rats. Parasitology. 1997 Jul; 115(1):9-14.

307 33. Jenkins MC, Parker C, Hill D, Pinckney RD, Dyer R, Dubey JP. *Neospora caninum* is
308 detected in feral rodents. Veterinary Parasitology. 2007 Jan 31; 143(2):161-5.

309
310
311
312
313
314
315
316
317
318

Preprint