<u>Original Article</u> Detection of Antibiotic Resistance Genes (*CTX-M*, *Van* A and *Van* B) of *Enterococcus faecalis* Isolated from Children with Bacteremia by RT-PCR

Sulaiman, A. M1*, Hussein, S. A2, Husain, V. I1

1. Techniques Department, College of Health and Medical Techniques Medical Lab, Northern Technical University, Mosul,

Iraq

2. Basic Sciences, College of Dentistry, the University of Sulaimani, Sulaymaniyah, Iraq

Received 25 June 2022; Accepted 10 July 2022 Corresponding Author: mostafa.master@gmail.com

Abstract

Fever is one of the most common diseases affecting humans, as it results from any disease or development and worsening of the disease for most people with widespread infections in the body. Therefore, this study aimed to evaluate antibiotic resistance genes (CTX-M, Van A and Van B) of Enterococcus faecalis isolated from children with bacteremia by RT-PCR. A total of 200 children was enrolled in the study, 100 children with fever and 100 healthy children (not suffering from any problem); that is, they are a control group for the detection of antibiotic resistance genes (CTX-M, Van A and Van B) of Enterococcus faecalis by RT-PCR. The age of the two groups ranged from one to five years. Four ml of venous blood sample was collected from each child; the venipuncture area was sterilized first with alcohol at a rate of 70%, followed by medical iodine and then sterilized with alcohol again to avoid contamination with skin flora. The blood samples were cultured on media for isolating bacteria. Then, the resistant isolates of E. faecalis to Vancomycin and cefotaxime antibiotics were taken and kept in special nutrient agar media where the DNA of the bacteria was extracted using (Zymogene Extraction kit, Japan). The detection of the exact genes (CTX-M, Van A and Van B) was done using Real-Time PCR technology according to the protocol mentioned by the company (Sacace biotechnology, Italy). The study presented that 40% of children with fever have positive blood cultures compared with 5% in the control group, with a significant difference between the two groups (P < 0.001). The study found that 32.5% of bacteremic children were due to S. aureus, 30%, 5%, and 4% were due to E. faecalis, E. coli, P. aeruginosa and Klebsiella spp, respectively, with significant difference (P < 0.01). The study showed that 91.67% of E. faecalis isolates were sensitive to Levofloxacin, 83.33% to Amoxiclav, 66.67% to Erythromycin, 58.33% to Amikacin, 50% to Ampicillin, 33.33% to cefotaxime and Ceftriaxone and 25% toward Vancomycin. From 9 isolates resistant to Vancomycin, the study presented that 88.89% of them were observed with Van A gene production as detected by real-time PCR (P<0.001). The study also showed that 77.78% were observed with Van B gene production as detected by real-time PCR (P<0.001). The study revealed that all E. faecalis isolates resistant to cefotaxime and Ceftriaxone were characterized by CTX gene production as detected by real-time PCR (P<0.001). Keywords: Antibiotic resistance, CTX-M, Vancomycin, Enterococcus faecalis RT-PCR

1. Introduction

Fever is one of the most common diseases affecting humans, as it results from any disease or development and worsening of the disease for most people with general infections in the body (1). Bacteremia in humans, especially children, is one of the most important diseases characterized by high temperatures as a result of the spread of many types of bacteria that are distinguished by their virulence and their ability to cause pathological events, given that they have many toxins and outputs that ultimately lead to high temperatures in these people (2). Among those bacteria, the Enterococcus faecalis is one of the most important causative bacteremia pathogens. Enterococcus faecalis is a gram-positive bacterium that causes several human infections (3). The problem of antibiotic resistance in bacteria isolated from sick people is one of the biggest problems facing medical personnel, children's burden and general surgeons because the increased rate of bacterial resistance to antibiotics makes them more dangerous as they lead and threaten the lives of patients at a very high rate, especially in societies where do not pay great attention to antibiotic resistance in bacteria (4). Several recent studies were conducted to identify and reveal genes that cause antibiotic resistance in many bacteria that cause bacteremia in children and adults worldwide (5-7). Therefore, this study aimed to evaluate antibiotic resistance genes (CTX-M, Van A & Van B) of Enterococcus faecalis isolated from children with bacteremia by RT-PCR.

2. Materials and Methods

2.1. Participants and Study Design

The study was conducted in the city of Kirkuk from April 2019 to January 2020 and included 100 children with fever and 100 healthy children not suffering from any problems as the control group. The ages of the two groups ranged from 1-5 years. The study included collecting blood samples from the children included in the study, where 4 ml of venous blood was collected from each child, where the withdrawal area was sterilized well with alcohol at a rate of 70%, followed by the use of medical iodine, and then sterilized with alcohol a second time to avoid contamination with skin flora.

2.2. Bacterial Isolation

Characterization of the isolated enterococci to the genus level was performed using Gram staining, blackening of Bile Aesculin Azide Agar (Oxoid), and culture on nutrient broth at 10°C, 45°C, with 6.5% NaCl. Then the motility test, sugar fermentation tests (L-Arabinose, Mannitol, Sorbitol Glycerol D-Lyxose Mannitol, Galactose, and Hippurate), and arginine dihydrolase and pyruvate utilization test were used for

characterization of the isolated enterococci to the species level.

2.3. Antimicrobial Susceptibility Testing

Antimicrobial susceptibility of the isolates was determined by disk diffusion method for the following antibacterial agents; Erythromycin (15 µg), Ampicillin (10 µg), Amoxiclave (30 µg), Levofloxacin (30 µg), Vancomycin (30 µg), Amikacin (30 µg), Cefotaxime (30 µg), Ceftriaxone (5 µg) (Bioanalyse, Turkey). Muller-Hinton agar plates were inoculated with 0.5 McFarland standard suspensions of the strains; antimicrobial disks were placed into plates and then incubated at 37°C for 24 h. The agar dilution method determined the minimum inhibitory concentrations (MICs) of Vancomycin and Erythromycin. Zone diameters were assessed according to the Clinical Laboratory Standard Institute guidelines.

2.4. DNA Extraction

Then, *E. faecalis* isolates, resistant to Vancomycin and Cefotaxime, were taken, isolated and kept in special nutrient agar media where the DNA of the bacteria was extracted using (Zymogene Extraction kit, Japan). Then, the exact genes (*CTX-M*, *Van A* and *Van B*) were detected using Real-Time PCR technology and according to the protocol mentioned by the company (Sacace biotechnology, Italy).

2.5. Statistical Analysis

Categorical variables were analyzed using the chisquare test using SPSS software (version 20). *P*-values of < .05 were considered statistically significant.

3. Results

The study presented that 40% of children with fever have positive blood cultures compared with 5% in the control group, with a significant difference between the two groups (P<0.001) (Table 1).

 Table 1. Prevalence of Bacteremia in children with and without fever

Decult	Children with fever		Healthy group	
Result	No.	%	No.	%
+ve blood culture	40	40	5	5
Negative	60	60	95	95
Total	100	100	100	100

P<0.001

The study found that 32.5% of bacteremic children were due to S. aureus, 30% to *E. faecalis*, 5% of *E. coli*, 4% of *P. aeruginosa* and *Klebsiella spp.* (*P*<0.01) (Table 2).

 Table 2. Distribution of bacterial isolates from bacteremic children

Bostorial isolatos	Children with fever		
Bacterial isolates	No.	%	
S. aureus	13	32.5	
E. faecalis	12	30	
E. coli	5	12.5	
P. aeruginosa	4	10	
Klebsiella spp	4	10	
S. epidermidis	2	5	
Total	40	100	

P<0.01

Table 3 shows that 91.67% of *E. faecalis* isolates are sensitive to Levofloxacin, 83.33% to Amoxiclave, 66.67% to Erythromycin, 58.33% to Amikacin, 50% to Ampicillin, 33.33% to Cefotaxime and Ceftriaxone and 25% toward Vancomycin.

 Table 3. Rate of antibiotics sensitivity toward isolated E.

 Faecalis

Antibiotics	Rate of antibiotic sensitivity (E. faecalis, n:12)		
Antibiotics	No.	%	
Ampicillin	6	50	
Levofloxacin	11	91.67	
Erythromycin	8	66.67	
Vancomycin	3	25	
Ceftriaxone	4	33.33	
Amikacin	7	58.33	
Amoxiclav	10	83.33	
Cefotaxime	4		

From 9 isolates which were resistant to Vancomycin, the study presented that 88.89% of them were observed with *Van A* gene production as detected by real-time PCR (P<0.001) (Table 4).

 Table 4. Rate of Van A gene production (by RT-PCR)

Van A gene	No.	%
Present	8	88.89
Absent	1	11.11
Total	9	100

P<0.001

The study also showed that 77.78% were observed with *Van B* gene production as detected by real-time PCR (P<0.001), as in table 5.

The study revealed that all *E. faecalis* isolated who were resistant to cefotaxime and Ceftriaxone were characterized by *CTX* gene production as detected by real-time PCR (P<0.001) (Table 6).

Table 5. Rate of Van B gene production (by RT-PCR)

Van B gene	No.	%
Present	7	77.78
Absent	2	11.11
Total	9	100

P<0.001

Table 6. Rate of CTX-M gene production (by RT-PCR)

CTX-M gene	No.	%
Present	8	100
Absent	0	0
Total	8	100

P<0.001

4. Discussion

The study presented that 40% of children with fever have positive blood cultures compared with 5% in the control group, with a significant difference between the two groups (P < 0.001). The results that arrived in our study were similar to the results of studies that were previously conducted in different countries of the world, as these studies confirmed that the most common causes of fever in children are bacteremia and sepsis, as the rates of bacteria isolated from blood samples with persons similar to the offending persons in our study reached a rate of up to to 45% in those studies (6, 7). The high temperature in people with bacteremia is due to the bacterial secretion of many toxins that lead to a high level of Interleukin 1 and gamma interferon, which leads to a rise in the temperature in the body (8). The study found that 32.5% of bacteremic children were due to S. aureus, 30% to E. faecalis, 5% of E. coli, 4% of P. aeruginosa and Klebsiella spp. (P<0.01), (Table 2). Table 3 shows that 91.67% of E. faecalis isolates are sensitive to 83.33% to Amoxiclave, 66.67% to Levofloxacin, Erythromycin, to Amikacin, 58.33% 50% to Ampicillin, 33.33% to Cefotaxime and Ceftriaxone and 25% toward Vancomycin. Several studies also found similar findings, as *S. aureus* and *E. faecalis* were the predominant bacterial isolates of bacteremic children (9, 10). Other studies indicated that most E. faecalis were resistant to Vancomycin (6, 8). Another study found that most E. faecalis isolates were resistant to Vancomycin and Ceftriaxone, with a rate reaching 80% (11). The reasons for the high resistance of these bacteria to many antibiotics are the fact that they are present and in abundance in the medical body as well as the community hospital environment (12) and that the excessive and wrong use of antibiotics in the community has a negative impact on these bacteria and made them resistant to antibiotics, which are used frequently and in excess in all pathological conditions such as urinary tract infection, diarrhea and coughing. On the other, by any means, it is more harmful than other types resistant to antibiotics (13, 14). It is worth noting that most of the types of antibiotic resistance genes in Enterococcus faecalis are the Van A and Van B gene and CTX B genes (15). As very recent studies reported that the highest percentage of antibioticresistance genes genetically isolated from bacteria that are resistant to antibiotics and that the most significant cause of antibiotic resistance in most types of bacteria, in addition to the bacteria mentioned above, is genetics, meaning that the reason is the transfer of genes from one bacterium to another through means like transformation and transduction (16-18).

Bacteria children were with Vancomycin and cephalosporin-resistant *E. faecalis* bacteria due to CTX-M, *Van A* and *Van B* production.

Authors' Contribution

Study concept and design: A. M. S. Acquisition of data: A. M. S. Analysis and interpretation of data: V. I. H. Drafting of the manuscript: S. A. H.

Critical revision of the manuscript for important intellectual content: S. A. H. Statistical analysis: V. I. H.

Administrative, technical, and material support: A. M. S.

Ethics

The study protocol was approved by the Northern Technical University, Mosul, Iraq ethics committee.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

- 1. Marston HD, Dixon DM, Knisely JM, Palmore TN, Fauci AS. Antimicrobial resistance. JAMA. 2016;316(11):1193-204.
- 2. Newland JG, Schuster J. Infections in Children, An Issue of Infectious Disease Clinics of North America, E-Book: Elsevier Health Sciences; 2018.
- 3. Friedman G, Stepensky P, Ahmad WA, Masarwa R, Temper V, Oster Y, et al. Enterococcal bacteremia in children with malignancies and following hematopoietic stem cell transplantation: a 15-year single-center experience. Pediatr Infect Dis J. 2020;39(4):318-24.
- 4. Morales-Espinosa R, Delgado G, Espinosa LF, Isselo D, Mendez JL, Rodriguez C, et al. Fingerprint analysis and identification of strains ST309 as a potential high risk clone in a Pseudomonas aeruginosa population isolated from children with bacteremia in Mexico City. Front Microbiol. 2017;8:313.
- 5. Beckman M, Washam MC, DeBurger B, Haslam DB, Courter JD, Andersen H, et al. Reliability of the Verigene system for the identification for Gram-positive Bacteria and detection of antimicrobial resistance markers from children with bacteremia. Diagn Microbiol Infect Dis. 2019;93(3):191-5.
- 6. Dodson DS, Dominguez SR, MacBrayne CE, Williams MC, Parker SK, editors. Vancomycinnonsusceptible enterococci mediated by vanC at a large children's hospital: prevalence, susceptibility, and impact on care of enterococcal bacteremia. Open Forum Infectious Diseases; 2020: Oxford University Press US.

- Furuichi M, Furuichi M, Horikoshi Y, Miyairi I. Infectious diseases consultation improves treatment and decreases mortality by enterococcal bacteremia in children. Pediatr Infect Dis J. 2018;37(9):856-60.
- Campbell AJ, Daley DA, Bell JM, Pang S, Coombs GW, Carapetis JR, et al. Progress towards a coordinated, national paediatric antimicrobial resistance surveillance programme: Staphylococcus aureus, enterococcal and Gram-negative bacteraemia in Australia. J Antimicrob Chemother. 2020;75(6):1639-44.
- 9. Kateete DP, Edolu M, Kigozi E, Kisukye J, Baluku H, Mwiine FN, et al. Species, antibiotic susceptibility profiles and van gene frequencies among enterococci isolated from patients at Mulago National Referral Hospital in Kampala, Uganda. BMC Infect Dis. 2019;19(1):1-9.
- 10. Rengaraj R, Mariappan S, Sekar U, Kamalanadhan of vancomycin resistance A. Detection among Enterococcus faecalis and Staphylococcus aureus. Journal diagnostic JCDR. of clinical and research: 2016;10(2):DC04.
- Shokoohizadeh L, Ekrami A, Labibzadeh M, Ali L, Alavi SM. Antimicrobial resistance patterns and virulence factors of enterococci isolates in hospitalized burn patients. BMC Res Notes. 2018;11(1):1-5.
- Hammerum AM, Baig S, Kamel Y, Roer L, Pinholt M, Gumpert H, et al. Emergence of vanA Enterococcus faecium in Denmark, 2005–15. J Antimicrob Chemother. 2017;72(8):2184-90.

- 13. Jahansepas A, Aghazadeh M, Rezaee MA, Hasani A, Sharifi Y, Aghazadeh T, et al. Occurrence of Enterococcus faecalis and Enterococcus faecium in various clinical infections: detection of their drug resistance and virulence determinants. Microb Drug Resist. 2018;24(1):76-82.
- 14. Raza T, Ullah SR, Mehmood K, Andleeb S. Vancomycin resistant Enterococci: A brief review. J Pak Med Assoc. 2018;68(5):768-72.
- 15. Sever JL, Ellenberg JH, Ley AC, Madden DL, Fuccillo DA, Tzan NR, et al. Toxoplasmosis: maternal and pediatric findings in 23,000 pregnancies. Pediatrics. 1988;82(2):181-92.
- 16. Almahdawy OT, Pricop R, Sadik O, Najee H, Pircalabioru GG, Marutescu L, et al. Description of vancomycin resistance genes in Enterococcus sp. clinical strains isolated from Bucharest, Romania. Rom Biotechnol Lett. 2019;24:395-9.
- 17. Guzman Prieto AM, van Schaik W, Rogers MR, Coque TM, Baquero F, Corander J, et al. Global emergence and dissemination of enterococci as nosocomial pathogens: attack of the clones? Front Microbiol. 2016;7:788.
- Hammerum AM, Justesen US, Pinholt M, Roer L, Kaya H, Worning P, et al. Surveillance of vancomycinresistant enterococci reveals shift in dominating clones and national spread of a vancomycin-variable vanA Enterococcus faecium ST1421-CT1134 clone, Denmark, 2015 to March 2019. Eurosurveillance. 2019;24(34):1900503.