Further Observations on the Susceptibility of Different Species of Lymnaea Snails of Iran to Miracidia of Fasciola hepatica and Fasciola gigantica

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Summary

Fascioliasis caused by Fasciola hepatica and F. gigantica is widely spread in Iran. Intermediate hosts of these parasites, lymnaea snails, have an important role in transmission of the disease to animals and human beings. The degree of susceptibility of limnaea of Iran to infection with F. hepatica and F. gigantica has been investigated and is reported in this paper.

Lymnaea truncatula and L. stagnalis act as intermediate hosts for F. hepatica, whereas L. Peregra, L. auricularia (gedrosiana), L. Palustris and L. Stagnalis act as intermediate hosts for F. gigantica, in most parts of Iran.

Introduction

Fascioliasis, caused by F. hepatica and F. gigantica, widely occurs in domestic animals in Iran. The disease in livestock and individual human beings has been reported by many workers. In 1989 an epidemic of human fascioliasis occurred in Iran, in the Caspian Sea region, where approximately 7000 of people were afflicted(1).

Lymnaea species, as intermediate hosts of Fasciolae, have crucial role in

completion of the life cycle and the transmission of the parasite to animals and man.

The common species of Lymnaea snails found in Iran are: L. truncatula, L. peregra, L. auricularia, L. palustris, L. stagnalis, L. auricularia (gedrosiana). Recently L. auricularia (rufescens) was reported to be present in south and south-east of Iran(2).

Some of Lymnaea snails, intermediate host for Fasciola spp in Iran, have been previously studied (3, 4, 5).

L. truncatula is omnipresent in the Iranian plateau and appears to be the most suitable intermediate host for F. hepatica in this country, as it is in Europe(6). In order to further study Lymnaea and determine the degree of susceptibility of Lymnaea spp, other than previously studied, to F. hepatica and F. gigantica miracidia, the experiments described in this paper were conducted.

Materials and methods

The aquatic snails, L. peregra, L. auricularia (gedrosiana), L. palustris were investigated. All Lymnaea used for the present experiments were reared in the snail chambers at Parasitology Department, Razi Institute. The origin of L. peregra and L. auricularia was Mardabad swamps, 60 km west of Tehran. They had been in culture at the laboratory since 1967. The origin of L. palustris was Babol and Babolsar rice fields in northern Iran. They had been cultured at our laboratory since 1978. The strain of L. stagnalis was from Uremia, a province in north-west of Iran.

The eggs were collected directly from the uterus of the worms F. hepatica and F. gigantica. Fasciolae were collected from bile ducts of sheep livers. These sheep, which were sacrificed, had been artificially infected with either F. hepatica or F. gigantica metacercariae.

The collected F. hepatica and F. gigantica eggs were cultured separately, in dark petri-dishes containing water, at 24° C for 10 to 15 days until development of miracidia, thenafter they were kept at 4° C in a refrigerator.

According to recommendations by Kendall et al.(7), and our previous experience, only juvenile snails were exposed to newly hatched miracidia, because snails are more susceptibile to infection at this particular age.

In order to determine the snails susceptibility to infection with miracidia of *Fasciolae*, the following experiments were conducted:

Experiment I: 120 L. peregra, 40-60 days old, were used. They were divided in three groups:

Group A: 40 snails were individually exposed to 1 newly hatched

- F. gigantica miracidium. Group B: 40 snails were individually exposed to 2 newly hatched
- F. giganticia miracidia. Gropu C: 40 snails were individually exposed to 2 newly hatched
- F. hepatica miracidia.

Experiment II: 124 L. auricularia (gedrosiana), were divided into three groups and were infected as follows:

Group A: 22 fifty-day old snails were exposed to 2 F. gigantica miracidia. Group B: 80 two- to four-week old snails were exposed to 1 single

F. gigantica miracidium.

Group C: 22 two- to four-week old snails were exposed to 2 F. hepatica miracidia.

Snails were, individually, exposed to miracidia for 4 h in a test tube containing small amount of pond water. Each group of snails was kept in a small, wide-mouth plastic container. They were daily checked for development of radiae and cercariae.

Experiment III: 865 L. palustris of different ages were exposed overnight to, approximately, 4 miracidia of either F. gigantica or F. hepatica miracidia as follows:

Group A: 250 adult snails were exposed to F. gigantica miracidia.

Group B: 255 juvenile, 1- to 3-week old, snails were also exposed to

F. gigantica miracidia.

Group C: 180 adult snail were exposed to F. hepatica miracidia.

Group D: 180 juvenile, 1- to 3-week old, were exposed to newly hatched miracidia of *F. hepatica*.

Each group of the exposed snails was kept in a snail room, in separate containers, and checked for development of rediae and cercariae.

Experiment IV: 360 adults and 360 juvenile L. stagnalis were exposed, according to the pattern described in Experiment III, to F. hepatica and

F. gigantica miracidia:

Group A: 240 adult snails were exposed to F. gigantica miracidia.

Group B: 240 juvenile snails, 1- to 3-week old, were exposed to newly hatched F. gigantica miracidia.

Group C: 120 adult snails were exposed to newly hatched F. hepatica miracidia.

Group D: The remaining 120 juvinile snails were exposed to F. hepatica miracidia.

Infected snails were kept, in separate containers, in a snail room and were checked daily for development of rediae and cercariae.

Results

L. peregra and L. auricularia (gedrosiana), in adult or immature stages, were very suitable intermediate hosts for F. gigantica miracidia and produced cercariae and metacercariae. Rates of susceptibility to infection, prepatant periods, from the time of exposure to miracidia till the first shedding of cercariae, number of cercariae produced when 1 or 2 miracidia were used are shwon in Table 1, for L. peregra, and in Table 2 for L. auricularia. Adult and juvenile L. peregra and L. auricularia (gedrosiana) were completely refractory to F. hepatica miracidia infection.

Rates of susceptibility to infection and prepatant periods, when 1 or 2 miracidia of F. gigantica were used, are shown in Table 3 for L. palustris. L. palustris, at adult stage, were completely refractory to F. gigantica and F. hepatica miracidia. They were also absolutely refractory to multiple exposure to F. hepatica miracidia. However, at juvenile stage, they were ussceptible to F. gigantica.

The adults of L. stagnalis were completely refractory to both F. hepatica and F. gigantica miracidia. On the contrary, the immatures of this snail were suscetible to both F. hepatica and F. gigantica. The results of the expetiment on this snail are summarised in Table 4. These results indicate that L. stagnalis is not a good intermediate host for animal fascioliasis.

Experimental subjects	F. gi	F. hepatica	
Lymnaea peregra number	Group A. 40	Group B. 40	Group C. 40
Snail age (days)	40-60	40-60	40-60
Exposed miracidia (number)	1	2	2
Snail infection rate	33.3%	86.6%	0%
Prepatent period for cercaria	70	57	0
Cercaria shedding period (days)	47	65	0
Mortality rate in prepatent period	32%	25%	0
Total cercaria excretion per snail	1971	3395	0
Mean life of infected Lymnaea (days)	117	124	0
Daily cercarial output per snail (Mean)	42	55	0

Table 1. Fasciola hepatica and Fasciola gigantica infection rates in Lymnaea peregra exposed, 4 h, to 1 or 2 miracidia.

Table 2. Observations on *L. auricularia* exposed, individually, in a test tube to 1 or 2 *F. gigantica* or *F. hepatica* miracidia for a period of 4 h.

Experimental subjects	L. auricularia (gedrosiana)				
Snail number	Group A. 22	Group B. 80	Group C. 22		
Lymnaea snail age	50	15-30	1 5-30		
Exposed miracidia number	2	1	2		
Fasciola species	F.gig.	F.hep.	F.hep.		
Snail infection rates	38%	80%	0%		
Prepatent period for cercaria (days)	70	56	0		
Mortality rate in prepatent period	27.2%	29%	5%		
Total excretion cercaria per snail	1164	1158	0		
Daily cercarial output per snail (number)	48	35	0		
Mean life-span of infected Lymnaea (days)	94	114	0		

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Experimental subjects	L. palustris				
	Group A. O	Group B.	Group C.	Group D.	
Snailnumber	250	255	180	180	
Snail age (weeks)	adult	1-3	adult	1-3	
Fasciola species	F.gig.	F.gig.	F.hep.	F.hep.	
Number of miracidia per snail	4	4	4	4	
prepatent period (days)	0	65	0	0	
Mortality rate in prepatent period	0	11%	0	0	
Infection rate	0	7 9%	0	0	
Daily mean cercaria shedding rate	0	17	0	0	
Mean life in infected snail (days)	0	89	0	0	

Table 3. Obser	vations on sus	ceptibility of	L. palustris	exposed, for 4 h,	to
one	or two miracio	lia of F. hepa	tica and F.	gigantica.	

Table 4. Susceptibility of L. stagnalis after exposure, for 4 h, to 4

Experimental subjects	L. stagnalis			
Experimental subjects	Group A.	Group B.	Group C.	Group D.
Miracidia source	F. gigantica		F. hepatica	
Snailnumber	240	240	120	120
Snail age (weeks)	adult	1-3	adult	1-3
Number of miracidia (per snail)	4	4	4	4
prepatent period (days)	0	58	0	71
Prepatent mortality	2%	22%	3%	79%
Infection rate in snails	0	75.8%	0	16.3%
Mean cercarial shedding per snail	0	11	0	47
Cercarial excerting period (days)	0	31	0	24

F. hepatica and F. gigantica.

Discussion

According to Massoud et al.(4) the infection rates of L. peregra and

L. auricularia to F. gigantica were 73% and 45%, respectively. In the present studies we almost found similar results. In addition, we also found that immature stages of the above mentioned snails were more susceptible to F. gigantica miracidia and showed 86.6% infection rate. Furthermore, the rates of mortality in infected snails were 32% to 25% in the prepatent period.

Arfaa et al.(3) reported that L. auricularia (gedrosiana) was susceptible to F. hepatica miracidia. However, our observations showed that adult and immature L. peregra and L. auricularia were completely refractory to infection with F. hepatica miracidia. Massoud et al.(4) reported that the source of Fasciolae eggs from which miracidia were obtained was from a naturally infected liver of a buffalo which, they suspected, must have carried a mixed infection with F. hepatica and

F. gigantica. On the basis of the present findings, we confirm the views of the latter authors and believe that Arfaa et al.(3) must had been dealing with F. gigantica.

Massoud et al.(4) reported that L. palustris was completely refractory to both F. hepatica and F. gigantica miracidia. In our study, we found that 1- to 3-week old L. palustris were susceptible to F. gigantica miracidia and produced cercaria and metacercaria which infected the animals, but were refractory to F. hepatica. We suspect that Massoud et al.(4) exposed adult snails which are completely refractory to both Fasciola species.

Our studies on *L. stagnalis* in Iran indicate that this species, which is abundant in north west and western parts of Iran, at the immature stage is susceptible to both *Fasciolae*. On the contrary, adults of *L. stagnalis* are completely refractory to both *F. hepatica* and *F. gigantica* miracidia (Table 4).

Moreover, as it is shown in Table 4 the number of cercariae shed by

L. stagnalis was limited because all infected snails died during the few cercarial shedding days. Therefore, it seems reasonable to believe that this species can not maintain the transmission of the infection in nature.

We conclude that L. truncatula and L. stagnalis act as the snail intermediate hosts for F. hepatica but L. peregra, L. auricularia (gedrosiana), L. palustris and L. stagnalis act as the intermediate host for

F. gigantica, in most parts of Iran.

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